Molecular Docking Studies of BCl2 towards Chronic Lymphocyctic Leukemia

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1. Abstract: Leukemia is a cancer that involves the blood-forming tissues of the bone marrow, spleen and lymph nodes. It is characterized by an uncontrolled production of abnormal, immature blood cells. When a person has leukemia, the bone marrow does not work properly. The bone marrow produces abnormal, immature cells, called leukemia cells. Bcl-2 also called as B cell lymphoma 2 is a protein encoded by BCL-2 genes in humans.Bcl-2 is an anti-apoptotic protein that inhibits apoptosis & thus it promotes cell growth. The most mutated gene in CLL is BCL-2. The current study aimed to identify high affinity small molecules which can block BCL-2 using Molecular Docking studies. Through Molecular Docking studies, Pubchem ID: 45028345, Survivin found to be the most effective compound bounding BCL-2 with a lowest rerank score of -168.022. Hence the same was used for the pharmacophore studies such as H-bond interaction, elector static interaction, Aromatic interaction, Hydrophilic interaction etc. Further studies can continue with Virtual screening, similarity searching for survivin inhibitor showed 312 compounds. The docking result of entire 312 virtual screened compounds showed PubChem CID101725649 found to be the most effective compound bounding BCL-2 with a lowest rerank score of -159.516.

Keywords: Molecular Docking, Virtual Screening, ADMET, BCL2, Chronic Lymphocytic Leukemia

2. Introduction

Cancer is uncontrolled division of cells that leads to malignancy,but another important reason of cancer is inhibition of programmed cell death.[1] Bcl-2 also called as B cell lymphoma 2 is a protein encoded by BCL-2 genes in humans.Bcl-2 is an anti-apoptotic protein that inhibits apoptosis & thus it promotes cell growth.It contains four BH domains,BH-4 domain is only present in anti-apoptotic proteins,not present in pro-apoptotic proteins that induces apoptosis.[2]

Bcl-2 protein induces cancers, as cancers are also caused by mutations in tumor suppressor genes like P53, over production of growth promoting factors & under production of growth inhibiting factors.[3] Bcl-2 protein belongs to a family of apoptosis regulatory gene products that may be antagonists of death or agonists of death. Over expression of Bcl-2 is observed in patients suffering from chronic lymphocyctic leukemia in which clonal propagation of B cells is seen in blood ,lymph nodes.[4]

Bcl-2 is located in outer membrane of mitochondria, nucleus & endoplasmic reticulum, it inhibits release of cytochrome c from mitochondria & inhibit caspases, a protease that induces apoptosis so that intrinsic apoptotic pathway is inhibited.[5] Expression of Bcl2 mRNA in chronic lymphocytic leukemia is high, therefore antisense RNA technological approaches have been used to down-regulate its expression.[6]

Micro-RNA named as miR-15a and miR-16-1negatively regulate the expression of antiapoptotic protein Bcl-2.[7] Bcl-2 protein is resistant to chemotherapy treatment also. Antisense oligonucleotide down-regulates the expression of Bcl-2 gene .[8]In the present study structure and ligand based approaches have been implemented to identify drug, which have better inhibitory potential for anti-apoptotic protein Bcl2.

3. Methodology

3.1 Selection of inhibitors:

In the present study, 57 established inhibitors of BCL2 that target CLL were selected. Some inhibitors were lacking 3D structures, so their 3D structures was created using Marvin Sketch and was saved in SDF format. Table[1] shows all those inhibitors that are having Pubchem ID with 3D structure and Pubchem ID with prepared 3D structure :

Table 1: Inhibitors selected for the study

Sl no	Inhibitors	Pubchem ID	Molecular weight(g/mol)	H- Bond donor	H-Bond acceptor	logpvalue	Ref
01	Venetoclax	49846579	868.447	3	11	8.2	[9]
02	Obatoclax mesylate	16727411	413.492	3	6		[10]
03	Navitoclax	24978538	974.611	2	14	9.6	[11]
04	Flavopiridol	5287969	401.843	3	6	3.3	[12]
05	7-hydroxy- staurosporine	72271	482.54	3	5	2.7	[12]
06	Ibrutinib	24821094	440.507	1	6	3.6	[13],[14]
07	Ofatumumab	6918251	242.238	1	6	-0.3	[14]
08	Survivin	45028345	1063.367	12	15	-1.4	[15]
09	MDM2 Inhibitor	10022508	377.972	2	3		[16]
10	SNS-032	3025986	380.525	2	7	3	[17]
11	Duvelisib	50905713	416.869	2	5	4.1	[18]
12	Fludarabine	657237	285.235	4	9	-0.6	[19]
13	Homoharringtonine	285033	545.629	2	10	0.8	[20]
14	Nutlin-3A	11433190	581.494	1	5	5.2	[21]
15	Isosorbide	12597	146.142	2	4	-1.4	[22]
16	Sabutoclax	46236925	700.788	8	8	8.9	[23]
17	Idelalisib	11625818	415.432	2	7	3.7	[24]
18	Antimycin A	14957	548.633	3	9	5.3	[25]
19	Salinomycin	3085092	751.011	4	11	5.7	[26]
20	Roscovitine	160355	354.458	3	6	3.2	[27]
21	Zolpidem	5732	307.397	0	2	2.5	[28]
22	Ha14-1	35490	409.236	1	7	3	[29]
23	Entinostat	4261	376.416	3	5	2	[30]
24	SGI-1776	24795070	405.425	1	8	4.3	[31]
25	Bendamustine	65628	358.263	1	4	2.9	[32]
26	Cyclophosphamide	2907	261.083	1	4	0.6	[33]
27	Dasatinib	3062316	488.007	3	9	3.6	[34]
28	Entinostat	4261	376.416				[35]
29	Romidepsin	121595947	540.694	4	8	2.2	[36]
30	Silvestrol	11787114	654.665	4	13	1.6	[37]
31	Salinosporamide A	11347535	313.778	2	4	1.8	[38]
32	Mitoxantrone	4212	444.488	8	10	1	[39]
33	JQ1 compound	46907787	456.989	0	0	4.9	[40]
34	Nucleozin	2863945	426.857	0	6	3.9	[41]

35	Gefitinib	123631	446.907	1	8	4.1	[42]
36	TW-37	11455910	573.704	4	6	7.8	[43]
37	Hyaluronidase	91820602	354.288	2	3		[44]
38	Midostaurin	9829523	570.649	1	4	4.8	[45]
39	Valproic Acid	3121	144.214	1	2	2.8	[46]
40	2-Naphthoxyacetic	8422	202.209	1	3	2.5	[47]
	acid						
41	Edaravone	4021	174.203	0	2	1.3	[47]
42	Wnt-C59	57519544	379.463	1	3	4.2	[48]
43	Frakefamide	5493563	563.63	6	8	1.4	[48]
44	Beta-Catenin	3463933	361.193	1	5	3.9	[48]
45	CPP32	1992	675.648	8	13	-1.3	[49]
46	Levomenthol	16666	156.269	1	1	3	[50]
47	Cephradine	38103	349.405	3	6	0.4	[50]
48	Chlorambucil	2708	304.211	1	3	1.7	[51],[54]
49	Lenalidomide	216326	259.265	2	4	-0.5	[52]
50	Pentostatin	439693	268.273	4	6	-2.1	[53]
51	Cladribine	20279	285.688	3	7	0.8	[54]
52	Doxorubicin	31703	543.525	6	12	1.3	[54]
53	Vincristine	5978	824.972	3	12	2.8	[55]
54	Prednisone	5865	358.434	2	5	1.5	[55]
55	GA101	91747743	520.813	0	6		[56]
56	Fostamatinib	11671467	580.466	4	15	1.6	[57]
57	Gossypol	3503	518.562	6	8	6.9	[58]

3.2 Retrieval of Ligand Structure, Optimization and Protein Preparation: The three dimensional structures of 57 BCL2 inhibitors were retrieved from NCBI's PubChem database. The structures of selected Bcl-2 inhibitors were optimized and cleaned in 3d format using Marvin View (MarvinView 5.6.0.2, 1998-2011, Copyright © ChemAxon Ltd) (Csizmadia, 2000).[59] Some inhibitors have no 3D structures available, so their 3D structures was made by using Marvin View. Further ligand preparation was done by taking the 3D structure of all those inhibitors embedded in LigPrep module of Schrodinger suite, 2013 (Schrodinger. LLC, New York, NY) and were optimized through OPLS 2005 force field algorithm.[60]The prepared ligands were saved in single SDF file for further docking studies.[61] The three-dimensional structure of Bcl-2 [PDB: 2XAO][62] was retrieved from the Protein Data Bank. The protein was prepared by removing all bound water molecules and ligands.

3.3 Molecular Docking studies:

Molecular docking program Molegro Virtual Docker (MVD) which incorporates highly efficient PLP (Piece wise Linear Potential) and MolDock scoring function provided a flexible docking platform.[63] The pre-prepared 57 ligands were saved in one single SDF file. PDB file of target protein consist preexisting ligands were removed and prepared by detecting cavities were found in which the first cavity bear highest volume and was targeted for further procedure of docking with ligands. The 3D structures of inhibitors (ligands) were docked in binding site of Bcl-2 receptor . Docking parameters were set to 0.20 Å as grid resolution, maximum iteration of 1500 and maximum population size of 50.

Energy minimization and hydrogen bonds were optimized after the docking. Simplex evolution was set at maximum steps of 300 with neighborhood distance factor of 1. Binding affinity and interactions of similar compounds with protein were evaluated on the basis of the internal ES (Internal electrostatic Interaction), internal hydrogen bond interactions and sp2-sp2 torsions. On the basis of MolDock rerank score, best interacting compound was selected from each class.

3.4 Virtual screening parameters:

On the basis of lowest rerank score of an established inhibitor obtained after filtered docking result, similarity searching was done against NCBI's Pubchem compound database to obtain best compound having greater affinity than this established inhibitor. The filtrations properties parameter set by component rule of Lipinski's rule of five at threshold $\geq 95\%$.

3.5 Drug – Drug comparative study: The unnamed complex structure was retrieved from established drug docking filtered result and was imported in Molecular docking program Molegro Virtual Docker (MVD). It was cleaned by removing all the ligands, constraints, and cavities except protein, then best posed drug with lowest rerank score was imported and exported as best drug docked file in PDB format. Then unnamed complex structure was retrieved from virtual screnning docking result and the procedure was repeated. The excel sheet was prepared to check all the affinities, hydrogen interaction, steric energy and lowest rerank score to indentify the best drug.

3.6 ADMET studies: ADMET studies of best established drug with pubchem CID-45028345 and best virtually screened drug with PubChem CID101725649 was carried out by admetSAR server.[64]

3.7 Pharmacophore Mapping: Pharmacophric mapping which involves ligand interactionpatterns, hydrogen bond interaction, hydrophobic interactions and solvent accessible surface area upon ligand binding was evaluated using Accelrys Discovery Studio 3.5 DS Visualizer.[65]

3.8 Softwares, Suites and Webservers used:

Pubchem database from NCBI was used to search and prepare library of similar chemical compounds for virtual screening. All the chemical structures were optimized in MarvinSketch 5.6.0.2, (1998-2011, Copyright © ChemAxon Ltd). Flexible Molecular docking of the compounds with target was completed using Molegro Virtual Docker 2010.4.0.0. Accelrys Discovery Studio® Visualizer 3.5.0.12158 (Copyright© 2005-12, Accelyrys Software Inc.) was used for visualization.

The ligands were optimized by using the software Schrodinger suite 2013 (Schrodinger.LLC, 2009, New York, NY). ADMET profiles were studied and calculated using admetSAR (Laboratory of Molecular Modeling and Design. Copyright© 2012 East China University of Science and Technology, Shanghai Key Laboratory for New Drug Drug Design). Accelrys Discovery Studio® Visualizer 3.5.0.12158 (Copyright© 2005-12, Accelyrys Software Inc.) was used for visualization.

4. Results and Discussions

4.1 Docking results

The docking studies of complete pre-established 57 drugs resulted as compound 08 is best established compound. Table [2] as the compound 08 is Survivin has pubchem CID-45028345 shows the higher affinity score direct toward our target protein and has the great affinity properties as molecular weight 1063.367 g/mol, hydrogen bond donor count 12 and hydrogen

bond acceptor count 15, topological polar surface area 385 A² and log value is -1.4. Thus the compound reveals the superior inhibitory affinity over protein BCL2.

S.No.	Ligand	Filename	MolDock Score	Rerank Score	Interaction	HBond
1	45028345	[00] 45028345.mol2	-261.513	-168.022	-241.93	-11.4443
2	45028345	[01] 45028345.mol2	-248.088	-151.08	-197.702	-9.59171
3	45028345	[02] 45028345.mol2	-231.855	-140.357	-200.256	-6.84983
4	1992	[02] 1992.mol2	-198.317	-133.767	-196.681	-8.24646
5	5493563	[00] 5493563.mol2	-164.718	-125.183	-179.608	-2.25351
6	1992	[03] 1992.mol2	-197.229	-124.922	-192.271	-6.09625
7	5493563	[04] 5493563.mol2	-152.809	-123.427	-184.456	-4.87982
8	24795070	[00] 24795070.mol2	-160.186	-122.675	-182.571	-1.79242
9	49846579	[00] 49846579.mol2	-182.686	-122.171	-216.697	-8.18314
10	5493563	[01] 5493563.mol2	-153.193	-118.845	-170.396	-2.88687

 Table 2: Docking scores of inhibitors arranged in descending order

4.2 Virtual Screening:

Further similarity searching for survivin inhibitor showed 312 compounds. Table [3] shows the docking result of entire 312 virtual screened compounds showed PubChem CID101725649 as a compound with high affinity. This compound has a molecular weight of 968.09 g/mol, 13 hydrogen bond donor, 18 hydrogen bond acceptor, a topological surface area of 430 A² and a log P value is -6. Suchwise, among all 312 compounds the drug with pubchem CID-101725649 has much potential inhibition against CLL cancer over the target protein BCL2.

Table 3: Virtual screened drugs docking result with reference to high affinity Survivin

S.No.	Ligand	Filename	MolDock Score	Rerank Score	Interaction	HBond
1	101725649	[00]101725649.mvdml	-228.959	-159.516	-231.403	-13.6032
2	101725649	[00]101725649.mvdml	-228.959	-159.516	-231.403	-13.6032
3	101725649	[00]101725649.mvdml	-228.959	-159.516	-231.403	-13.6032
4	101725649	[00]101725649.mvdml	-228.959	-159.516	-231.403	-13.6032
5	23496768	[00]23496768.mvdml	-227.357	-159.013	-211.57	-10.7869
6	23496768	[00]23496768.mvdml	-227.357	-159.013	-211.57	-10.7869
7	23496768	[00]23496768.mvdml	-227.357	-159.013	-211.57	-10.7869
8	23496768	[00]23496768.mvdml	-227.357	-159.013	-211.57	-10.7869
9	101129057	[01]101129057.mvdml	-214.502	-158.611	-224.738	-8.41583
10	101129057	[01]101129057.mvdml	-214.502	-158.611	-224.738	-8.41583

4.3 Drug – Drug comparative study:

Table [4] shows the rerank scores of the established drug & virtually screened drug against the target protein BCL2 on CLL cancer. Hence, total energy of established inhibitor with pubchem CID 45028345 is lowest among the entire virtual screened compound with preferable affinity. Surprisingly, the other interaction of both the compounds displaying the virtual screened compound with pubchem CID 101725649 has more affinity interaction properties according to steric energy of PLP (Piecewise Linear Potential) and steric energy of LJ12-6(Leonard-Jones approximation) than the established drug, whereas the stability of hydrogen bonds is more in virtually screened drug than in established drug. So it demonstrates that the both the compounds have equivalent potential inhibition towards the target protein BCL2.

	Established		Virtual screen	ed
Energy	MolDock	Rerank	MolDock	Rerank
overview:Descriptors	Score	Score	Score	Score
Total Energy	-263.842	-169.88	-229.497	-159.935
External Ligand				
interactions	-244.255	-202.261	-231.938	-196.908
Protein - Ligand				
interactions	-244.255	-202.261	-231.938	-196.908
Steric (by PLP)	-196.256	-134.632	-217.797	-149.408
Steric (by LJ12-6)		-33.008		-36.299
Hydrogen bonds	-13.758	-10.896	-14.141	-11.2
Hydrogen bonds (no direction	onality)	0		0
Electrostatic (short range)	-24.978	-22.28	0	0
Electrostatic (long range)	-9.263	-1.445	0	0
Cofactor - Ligand	0	0	0	0
Steric (by PLP)	0		0	
Steric (by LJ12-6)		0		0
Hydrogen bonds	0	0	0	0
Electrostatic	0	0	0	0
Water - Ligand interactions	0	0	0	0
Internal Ligand				
interactions	-19.587	32.381	2.441	36.973
Torsional strain	36.26	34.012	31.332	29.39
Torsional strain (sp2-sp2)		0		0
Hydrogen bonds		0		0
Steric (by PLP)	-35.873	-6.17	-2.395	-0.412
. , ,				

Table 4: Drug – Drug comparative study

Juni Khyat (UGC Care Group I Listed	l Journal)	Vol-10 Is	ISSN: 2 ssue-6 No. 13 J	278-4632 June 2020	
Steric (by LJ12-6)		5.081		7.995	
Electrostatic	-1.242	-0.543	0	0	
Soft Constraint Penalty	0		0		
Search Space Penalty	0		0		

4.4 ADMET studies:

Table 5: ADMET profiles of Established drug and Virtual screened compound by AdmetSAR

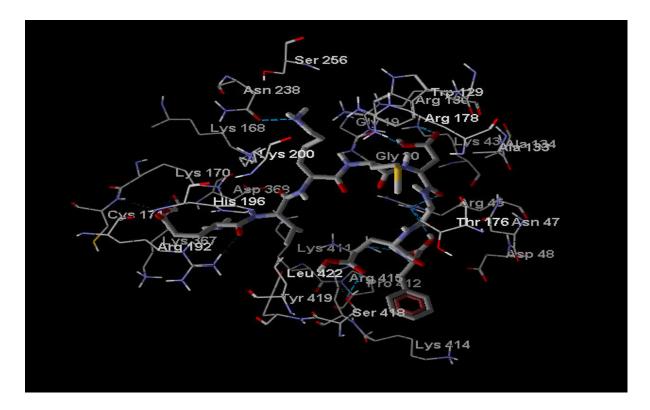
Properties		Established drug	Virtual screened compound
Absorption Probability	Result	Probability	•
Blood-Brain Barrier	BBB-	0.7522	0.8447
Human Intestinal Absorption	HIA+	0.8327	0.6152
Caco-2 Permeability	Caco2-	0.7121	0.7117
P-glycoprotein Substrate	Substrate	0.8212	0.8203
	Non-inhibitor	0.8558	0.8614
P-glycoprotein Inhibitor Renal Organic Cation	Non-inhibitor	0.9927	0.9903
Transporter	Non-inhibitor	0.9293	0.9443
Distribution	M ² (, , 1, , , , 1, 1, .	0.(200	0.5405
Subcellular localization Metabolism	Mitochondria	0.6209	0.5485
CYP450 2C9 Substrate	Non-substrate	0.8547	0.8138
CYP450 2D6 Substrate	Non-substrate	0.7651	0.738
CYP450 3A4 Substrate	Non-substrate	0.5806	0.5759
CYP450 1A2 Inhibitor	Non-inhibitor	0.9335	0.8983
CYP450 2C9 Inhibitor	Non-inhibitor	0.8844	0.8843
CYP450 2D6 Inhibitor	Non-inhibitor	0.8957	0.9093
CYP450 2C19 Inhibitor	Non-inhibitor	0.8406	0.8509
CYP450 3A4 Inhibitor	Non-inhibitor Low CYP Inhibitory	0.9061	0.9585
CYP Inhibitory Promiscuity	Promiscuity	0.9936	0.9894
Excretion Toxicity			
Human Ether-a-go-go-Related	Weak inhibitor	0.9836	0.986
Gene Inhibition	Non-inhibitor Non AMES	0.8819	0.8398
AMES Toxicity	toxic Non-	0.855	0.8535
Carcinogens	carcinogens	0.9342	0.9386
Fish Toxicity	High FHMT	0.9575	0.9171

Tetrahymena Pyriformis			
Toxicity	High TPT	0.9853	0.9561
Honey Bee Toxicity	Low HBT Not ready	0.7953	0.7802
Biodegradation	biodegradable	0.9476	0.9266
Acute Oral Toxicity	III	0.7257	0.6933
Carcinogenicity (Three-class)	Non-required	0.7008	0.7195

ADMET Predicted Profile Regression

			Established drug	Virtual drug	screened
	Model	Unit	Values	Values	
Absorption					
	Aqueous solubility	LogS	-3.1573	-2.6622	
	Caco-2 Permeability	LogPapp, cm/s	0.5011	0.2829	
Distribution					
Metabolism					
Excretion					
Toxicity					
	Rat Acute Toxicity	LD50, mol/kg	2.1486	2.2399	
	Fish Toxicity	pLC50, mg/L	1.8413	1.9241	
	Tetrahymena Pyriformis Toxicity	pIGC50, ug/L	0.2731	0.2037	

4.5 Pharmacophore Study:



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Most effective Virtual screened compound(PubID: 101725649) showing H-Bond interactions

5. Conclusion: Inhibition of BCl2 protein has now been as an important target of drug against Chronic Lymphocyctic Leukemia. Established drug Survivin with pubchem CID-45028345 & virtually screened drug with pubchem CID-101725649 are established inhibitors which are in clinical trials and promise to be potent drug in the near future. Our study utilizing in silico approaches has indicated that compounds 45028345 and 101725649 had better inhibitory potential for BCL2 anti-apoptotic protein.

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7. References

- 1. Cory, S., & Adams, J. M. (2002). The Bcl2 family: regulators of the cellular life-or-death switch. *Nature Reviews Cancer*, 2(9), 647.
- Khemtémourian, L., Sani, M. A., Bathany, K., Gröbner, G., & Dufourc, E. J. (2006). Synthesis and secondary structure in membranes of the Bcl-2 anti-apoptotic domain BH4. *Journal of Peptide Science*, 12(1), 58-64.
- **3**. Klobusicka, M., Kusenda, J., & Babusikova, O. (2002). Immunocytochemical detection of bcl-2 and p53 proteins in B-chronic lymphocytic leukemia patients. *Neoplasma*, 49(6), 387-393.
- Mariano, M. T., Moretti, L., Donelli, A., Grantini, M., Montagnani, G., Di Prisco, A. U., ... & Narni, F. (1992). bcl-2 gene expression in hematopoietic cell differentiation. *Blood*, 80(3), 768-775..
- Garrido, C., Galluzzi, L., Brunet, M., Puig, P. E., Didelot, C., & Kroemer, G. (2006). Mechanisms of cytochrome c release from mitochondria. *Cell death and differentiation*, 13(9), 1423.
- 6. Oltersdorf, T., Elmore, S. W., Shoemaker, A. R., Armstrong, R. C., Augeri, D. J., Belli, B. A., ... & Joseph, M. K. (2005). An inhibitor of Bcl-2 family proteins induces regression of solid tumours. *Nature*, 435(7042), 677.

- Cimmino, A., Calin, G. A., Fabbri, M., Iorio, M. V., Ferracin, M., Shimizu, M., ... & Rassenti, L. (2005). miR-15 and miR-16 induce apoptosis by targeting BCL2. Proceedings of the National Academy of Sciences of the United States of America, 102(39), 13944-13949.
- 8. O'brien, S. M., Cunningham, C. C., Golenkov, A. K., Turkina, A. G., Novick, S. C., & Rai, K. R. (2005). Phase I to II multicenter study of oblimersen sodium, a Bcl-2 antisense oligonucleotide, in patients with advanced chronic lymphocytic leukemia. *Journal of Clinical Oncology*, 23(30), 7697-7702.
- 9. Del Poeta, G., Postorino, M., Pupo, L., Del Principe, M. I., Dal Bo, M., Bittolo, T., ...&Venditti, A. (2016). Venetoclax: Bcl-2 inhibition for the treatment of chronic lymphocytic leukemia. *Drugs of today (Barcelona, Spain: 1998)*, 52(4), 249-260.
- O'Brien, S. M., Claxton, D. F., Crump, M., Faderl, S., Kipps, T., Keating, M. J., ... & Cheson, B. D. (2009). Phase I study of obatoclax mesylate (GX15-070), a small molecule pan–Bcl-2 family antagonist, in patients with advanced chronic lymphocytic leukemia. *Blood*, 113(2), 299-305.
- Roberts, A. W., Seymour, J. F., Brown, J. R., Wierda, W. G., Kipps, T. J., Khaw, S. L., ... & Cui, Y. (2011). Substantial susceptibility of chronic lymphocytic leukemia to BCL2 inhibition: results of a phase I study of navitoclax in patients with relapsed or refractory disease. *Journal of Clinical Oncology*, *30*(5), 488-496.
- 12. Kitada, S., Zapata, J. M., Andreeff, M., & Reed, J. C. (2000). Protein kinase inhibitors flavopiridol and 7-hydroxy-staurosporine down-regulate antiapoptosis proteins in B-cell chronic lymphocytic leukemia. *Blood*, *96*(2), 393-397.
- 13. Byrd, J. C., Furman, R. R., Coutre, S. E., Flinn, I. W., Burger, J. A., Blum, K. A., ... & Jones, J. A. (2013). Targeting BTK with ibrutinib in relapsed chronic lymphocytic leukemia. *New England Journal of Medicine*, *369*(1), 32-42.
- Byrd, J. C., Brown, J. R., O'brien, S., Barrientos, J. C., Kay, N. E., Reddy, N. M., ... & Devereux, S. (2014). Ibrutinib versus of atumumab in previously treated chronic lymphoid leukemia. *New England Journal of Medicine*, 371(3), 213-223.

- 15. Granziero, L., Ghia, P., Circosta, P., Gottardi, D., Strola, G., Geuna, M., ... & Caligaris-Cappio, F. (2001). Survivin is expressed on CD40 stimulation and interfaces proliferation and apoptosis in B-cell chronic lymphocytic leukemia. *Blood*, 97(9), 2777-2783.
- 16. Coll-Mulet, L., Iglesias-Serret, D., Santidrián, A. F., Cosialls, A. M., de Frias, M., Castaño, E., ... & Vassilev, L. T. (2006). MDM2 antagonists activate p53 and synergize with genotoxic drugs in B-cell chronic lymphocytic leukemia cells. *Blood*, 107(10), 4109-4114.
- 17. Walsby, E., Lazenby, M., Pepper, C., & Burnett, A. K. (2011). The cyclin-dependent kinase inhibitor SNS-032 has single agent activity in AML cells and is highly synergistic with cytarabine. *Leukemia*, 25(3), 411.
- Patel, V. M., Balakrishnan, K., Guerrieri, R., Wierda, W., O'Brien, S., & Gandhi, V. (2015). Elevated level of BCL-2 is the primary target for inhibition during duvelisib (IPI-145) therapy: ABT-199 neutralizes the resistance mechanism in chronic lymphocytic leukemia.
- 19. Walsby, E., Pratt, G., Shao, H., Abbas, A. Y., Fischer, P. M., Bradshaw, T. D., ... & Pepper, C. (2014). A novel Cdk9 inhibitor preferentially targets tumor cells and synergizes with fludarabine. *Oncotarget*, 5(2), 375.
- 20. Chen, R., Guo, L., Chen, Y., Jiang, Y., Wierda, W. G., & Plunkett, W. (2011). Homoharringtonine reduced Mcl-1 expression and induced apoptosis in chronic lymphocytic leukemia. *Blood*, 117(1), 156-164.
- 21. Kojima, K., Konopleva, M., McQueen, T., O'Brien, S., Plunkett, W., & Andreeff, M. (2006). Mdm2 inhibitor Nutlin-3a induces p53-mediated apoptosis by transcription-dependent and transcription-independent mechanisms and may overcome Atm-mediated resistance to fludarabine in chronic lymphocytic leukemia. *Blood*, 108(3), 993-1000.
- Anderson, M. A., Huang, D., & Roberts, A. (2014, July). Targeting BCL2 for the treatment of lymphoid malignancies. In *Seminars in hematology* (Vol. 51, No. 3, pp. 219-227). WB Saunders.
- 23. Pan, R., Ruvolo, V. R., Wei, J., Konopleva, M., Reed, J. C., Pellecchia, M., ... & Ruvolo, P. P. (2015). Inhibition of Mcl-1 with the pan–Bcl-2 family inhibitor (–) BI97D6 overcomes ABT-737 resistance in acute myeloid leukemia. *Blood*, *126*(3), 363-372.

- Brown, J. R., Byrd, J. C., Coutre, S. E., Benson, D. M., Flinn, I. W., Wagner-Johnston, N. D., ... & Johnson, D. M. (2014). Idelalisib, an inhibitor of phosphatidylinositol 3-kinase p110δ, for relapsed/refractory chronic lymphocytic leukemia. *Blood*, *123*(22), 3390-3397.
- Campàs, C., Cosialls, A. M., Barragán, M., Iglesias-Serret, D., Santidrián, A. F., Coll-Mulet, L., ... & Gil, J. (2006). Bcl-2 inhibitors induce apoptosis in chronic lymphocytic leukemia cells. *Experimental hematology*, 34(12), 1663-1669.
- 26. Lu, D., Choi, M. Y., Yu, J., Castro, J. E., Kipps, T. J., & Carson, D. A. (2011). Salinomycin inhibits Wnt signaling and selectively induces apoptosis in chronic lymphocytic leukemia cells. *Proceedings of the National Academy of Sciences*, 108(32), 13253-13257.
- 27. Hahntow, I. N., Schneller, F., Oelsner, M., Weick, K., Ringshausen, I., Fend, F., ... & Decker, T. (2004). Cyclin-dependent kinase inhibitor Roscovitine induces apoptosis in chronic lymphocytic leukemia cells. *Leukemia*, *18*(4), 747.
- 28. Cimmino, A., Calin, G. A., Fabbri, M., Iorio, M. V., Ferracin, M., Shimizu, M., ... & Rassenti, L. (2005). miR-15 and miR-16 induce apoptosis by targeting BCL2. *Proceedings of the National Academy of Sciences of the United States of America*, 102(39), 13944-13949.
- Campàs, C., Cosialls, A. M., Barragán, M., Iglesias-Serret, D., Santidrián, A. F., Coll-Mulet, L., ... & Gil, J. (2006). Bcl-2 inhibitors induce apoptosis in chronic lymphocytic leukemia cells. *Experimental hematology*, 34(12), 1663-1669.
- Lucas, D. M., Davis, M. E., Parthun, M. R., Mone, A. P., Kitada, S., Cunningham, K. D., ... & Grever, M. R. (2004). The histone deacetylase inhibitor MS-275 induces caspasedependent apoptosis in B-cell chronic lymphocytic leukemia cells. *Leukemia*, 18(7), 1207.
- Chen, L. S., Redkar, S., Bearss, D., Wierda, W. G., & Gandhi, V. (2009). Pim kinase inhibitor, SGI-1776, induces apoptosis in chronic lymphocytic leukemia cells. *Blood*, *114*(19), 4150-4157.
- 32. Hallek, M. (2013). Signaling the end of chronic lymphocytic leukemia: new frontline treatment strategies. *Blood*, *122*(23), 3723-3734.
- Eichhorst, B., Robak, T., Montserrat, E., Ghia, P., Hillmen, P., Hallek, M., & Buske, C. (2015). Chronic lymphocytic leukaemia: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Annals of Oncology*, 26(suppl_5), v78-v84.

- 34. Veldurthy, A., Patz, M., Hagist, S., Pallasch, C. P., Wendtner, C. M., Hallek, M., & Krause, G. (2008). The kinase inhibitor dasatinib induces apoptosis in chronic lymphocytic leukemia cells in vitro with preference for a subgroup of patients with unmutated IgV H genes. *Blood*, 112(4), 1443-1452.
- 35. Lucas, D. M., Davis, M. E., Parthun, M. R., Mone, A. P., Kitada, S., Cunningham, K. D., ... & Grever, M. R. (2004). The histone deacetylase inhibitor MS-275 induces caspasedependent apoptosis in B-cell chronic lymphocytic leukemia cells. *Leukemia*, 18(7), 1207.
- 36. Oda, E., Ohki, R., Murasawa, H., Nemoto, J., Shibue, T., Yamashita, T., ... & Tanaka, N. (2000). Noxa, a BH3-only member of the Bcl-2 family and candidate mediator of p53-induced apoptosis. *Science*, 288(5468), 1053-1058.
- 37. Lucas, D. M., Edwards, R. B., Lozanski, G., West, D. A., Shin, J. D., Vargo, M. A., ... & Goettl, V. M. (2009). The novel plant-derived agent silvestrol has B-cell selective activity in chronic lymphocytic leukemia and acute lymphoblastic leukemia in vitro and in vivo. *Blood*, *113*(19), 4656-4666.
- 38. Ruiz, S., Krupnik, Y., Keating, M., Chandra, J., Palladino, M., & McConkey, D. (2006). The proteasome inhibitor NPI-0052 is a more effective inducer of apoptosis than bortezomib in lymphocytes from patients with chronic lymphocytic leukemia. *Molecular Cancer Therapeutics*, 5(7), 1836-1843.
- 39. Bellosillo, B., Colomer, D., Pons, G., & Gil, J. (1998). Mitoxantrone, a topoisomerase II inhibitor, induces apoptosis of B-chronic lymphocytic leukaemia cells. *British journal of haematology*, *100*(1), 142-146.
- Ott, C. J., Kopp, N., Bird, L., Paranal, R. M., Qi, J., Bowman, T., ... & Weinstock, D. M. (2012). BET bromodomain inhibition targets both c-Myc and IL7R in high-risk acute lymphoblastic leukemia. *Blood*, *120*(14), 2843-2852.
- Otake, Y., Soundararajan, S., Sengupta, T. K., Kio, E. A., Smith, J. C., Pineda-Roman, M., ... & Fernandes, D. J. (2007). Overexpression of nucleolin in chronic lymphocytic leukemia cells induces stabilization of bcl2 mRNA. *Blood*, *109*(7), 3069-3075.
- 42. Iorio, M. V., & Croce, C. M. (2009). MicroRNAs in cancer: small molecules with a huge impact. *Journal of clinical oncology*, 27(34), 5848-5856.

- 43. Mohammad, R. M., Goustin, A. S., Aboukameel, A., Chen, B., Banerjee, S., Wang, G., ... & Al-Katib, A. (2007). Preclinical studies of TW-37, a new nonpeptidic small-molecule inhibitor of Bcl-2, in diffuse large cell lymphoma xenograft model reveal drug action on both Bcl-2 and Mcl-1. *Clinical Cancer Research*, *13*(7), 2226-2235.
- 44. Shanafelt, T. D., Call, T. G., Zent, C. S., LaPlant, B., Bowen, D. A., Roos, M., ... & Yang, C. S. (2009). Phase I trial of daily oral Polyphenon E in patients with asymptomatic Rai stage 0 to II chronic lymphocytic leukemia. *Journal of Clinical Oncology*, 27(23), 3808-3814.
- 45. Ganeshaguru, K., Wickremasinghe, R. G., Jones, D. T., Gordon, M., Hart, S. M., Virchis, A. E., ... & Csermak, K. (2002). Actions of the selective protein kinase C inhibitor PKC412 on B-chronic lymphocytic leukemia cells in vitro. *haematologica*, 87(2), 167-176.
- 46. Bokelmann, I., & Mahlknecht, U. (2008). Valproic acid sensitizes chronic lymphocytic leukemia cells to apoptosis and restores the balance between pro-and antiapoptotic proteins. *Molecular Medicine*, *14*(1-2), 20.
- 47. Smit, L. A., Hallaert, D. Y., Spijker, R., de Goeij, B., Jaspers, A., Kater, A. P., ... & Eldering, E. (2007). Differential Noxa/Mcl-1 balance in peripheral versus lymph node chronic lymphocytic leukemia cells correlates with survival capacity. *Blood*, 109(4), 1660-1668.
- 48. Gandhirajan, R. K., Staib, P. A., Minke, K., Gehrke, I., Plickert, G., Schlösser, A., ... & Kreuzer, K. A. (2010). Small molecule inhibitors of Wnt/β-catenin/lef-1 signaling induces apoptosis in chronic lymphocytic leukemia cells in vitro and in vivo. *Neoplasia*, *12*(4), 326IN4-335IN6.
- Krajewski, S., Gascoyne, R. D., Zapata, J. M., Krajewska, M., Kitada, S., Chhanabhai, M., ... & Liu, Y. J. (1997). Immunolocalization of the ICE/Ced-3–Family Protease, CPP32 (Caspase-3), in Non-Hodgkin9s Lymphomas, Chronic Lymphocytic Leukemias, and Reactive Lymph Nodes. *Blood*, 89(10), 3817-3825.
- Bellosillo, B., Dalmau, M., Colomer, D., & Gil, J. (1997). Involvement of CED-3/ICE proteases in the apoptosis of B-chronic lymphocytic leukemia cells. *Blood*, 89(9), 3378-3384.
- 51. Hallek, M. (2015). Chronic lymphocytic leukemia: 2015 update on diagnosis, risk stratification, and treatment. *American journal of hematology*, *90*(5), 446-460.

- 52. Herman, S. E., Lapalombella, R., Gordon, A. L., Ramanunni, A., Blum, K. A., Jones, J., ... & Byrd, J. C. (2011). The role of phosphatidylinositol 3-kinase-δ in the immunomodulatory effects of lenalidomide in chronic lymphocytic leukemia. *Blood*, *117*(16), 4323-4327.
- 53. Rozman, C., & Montserrat, E. (1995). Chronic lymphocytic leukemia. *New England Journal of Medicine*, *333*(16), 1052-1057.
- 54. Bosanquet, A. G., Sturm, I., Wieder, T., Essmann, F., Bosanquet, M. I., Head, D. J., ... & Daniel, P. T. (2002). Bax expression correlates with cellular drug sensitivity to doxorubicin, cyclophosphamide and chlorambucil but not fludarabine, cladribine or corticosteroids in B cell chronic lymphocytic leukemia. *Leukemia*, 16(6), 1035.
- Stankovic, T., Weber, P., Stewart, G., Bedenham, T., Murray, J., Byrd, P. J., ... & Taylor, A. M. R. (1999). Inactivation of ataxia telangiectasia mutated gene in B-cell chronic lymphocytic leukaemia. *The Lancet*, 353(9146), 26-29.
- 56. Alduaij, W., Ivanov, A., Honeychurch, J., Cheadle, E. J., Potluri, S., Lim, S. H., ... & Glennie, M. J. (2011). Novel type II anti-CD20 monoclonal antibody (GA101) evokes homotypic adhesion and actin-dependent, lysosome-mediated cell death in B-cell malignancies. *Blood*, *117*(17), 4519-4529.
- 57. Hallek, M. (2013). Chronic lymphocytic leukemia: 2013 update on diagnosis, risk stratification and treatment. *American journal of hematology*, *88*(9), 803-816.
- Balakrishnan, K., Wierda, W. G., Keating, M. J., & Gandhi, V. (2008). Gossypol, a BH3 mimetic, induces apoptosis in chronic lymphocytic leukemia cells. *Blood*, *112*(5), 1971-1980.
- 59. Vuree, S., Dunna, N. R., Khan, I. A., Alharbi, K. K., Vishnupriya, S., Soni, D., ... & Nayarisseri, A. (2013). Pharmacogenomics of drug resistance in Breast Cancer Resistance Protein (BCRP) and its mutated variants. *Journal of Pharmacy Research*, 6(7), 791-798.
- 60. Jorgensen, W. L., Maxwell, D. S., & Tirado-Rives, J. (1996). Development and testing of the OPLS all-atom force field on conformational energetics and properties of organic liquids. *Journal of the American Chemical Society*, *118*(45), 11225-11236

- 61. Bandaru, S., Ponnala, D., Lakkaraju, C., Kumar, C., Bhukya, U. S., & Nayarisseri, A. (2014). Identification of high affinity non-peptidic small molecule inhibitors of MDM2p53 interactions through structure-based virtual screening strategies. *Asian Pac J Cancer Prev*, 16, 3759-65.
- 62. González, B., Baños-Sanz, J. I., Villate, M., Brearley, C. A., & Sanz-Aparicio, J. (2010). Inositol 1, 3, 4, 5, 6-pentakisphosphate 2-kinase is a distant IPK member with a singular inositide binding site for axial 2-OH recognition. *Proceedings of the National Academy* of Sciences, 107(21), 9608-9613.
- 63. Bandaru, S., Gangadharan Sumithnath, T., Sharda, S., Lakhotia, S., Sharma, A., Jain, A., ... & Kumar Singh, S. (2017). Helix-Coil Transition Signatures B-Raf V600E Mutation and Virtual Screening for Inhibitors Directed Against Mutant B-Raf. *Current drug metabolism*, 18(6), 527-534.
- 64. Cheng F, Li W, Zhou Y, et al (2012). AdmetSAR: a comprehensivesource and free tool for assessment of chemical ADMETproperties. J Chem Inf Model, 52, 3099-105.
- 65. Patidar, K., Deshmukh, A., Bandaru, S., Lakkaraju, C., Girdhar, A., Vr, G., ... & Singh, S. K. (2016). Virtual Screening Approaches in Identification of Bioactive Compounds Akin to Delphinidin as Potential HER2 Inhibitors for the Treatment of Breast Cancer. *Asian Pacific Journal of Cancer Prevention*, 17(4), 2291-2295.